

N-alkylation methods, Characterization and Evaluation of antibacterial activity of some Novel 5-Chloroisatin Derivatives

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ABSTRACT:

A series of new 5-Chloroisatin derivatives have been synthesized by the method of N-alkylation at room temperature, in the presence of a base and a catalyst with good yields. The chemical structures of these compounds were confirmed by NMR (¹H & ¹³C), these new compounds obtained were evaluated for their antibacterial activity. The final results revealed that the majority of the compounds exhibited good antimicrobial activity against various organisms.

Keywords: 5-Chloroisatin derivatives, N-alkylation, antibacterial activity, NMR

I. INTRODUCTION

The design, synthesis and production of molecules, are ones of the important objectives of organic and medicinal chemistry [1], which are having highly therapeutic interest. Since the discovery of heterocyclic nucleus the chemistry of isatin and their fused derivatives has a wide variety of pharmacological activities such as antimicrobial, anticancer, antiviral, anticonvulsant, anti-inflammatory and analgesic [2]. Different research group attempted study on isatin synthetic aspect. Other research group attempted study of isatin biological activity [3].

Thus the structural modification of the isatin derivatives moiety is still of the highest importance. This article has focused on the application of synthetic N-alkylation methods in the conditions of charge transfer catalysis in the presence of a base (K₂CO₃) and a catalyst (BTBA) between 5-Chloro-1H-indole-2,3-dione and alkylating agents with good to excellent yields [4]. The synthesized 5-Chloroisatin derivatives were characterized by spectroscopic techniques, such as, ¹H NMR, ¹³C NMR. In addition, they were evaluated for their biological activities [5] as antibacterial in vitro against two Gram positive bacteria such as *Staphylococcus aureus*, *Bacillus cereus* and two Gram negative bacteria such as *Escherichia coli*, *pseudomonas aeruginosa*.

II. MATERIAL AND METHODS

1. Chemistry:

All melting points were measured by the Kofler bench. The ¹H NMR, ¹³C NMR spectra were recorded on Bruker 300 NMR spectrometer advancement in CDCl₃ using tetramethylsilane (TMS) as reference. Chemical shifts are reported in parts per million. Reactions were monitored by thin layer chromatography (TLC) on silica gel, plates were visualizing with ultraviolet light or iodine. Column chromatography was performed on silica gel 60 (0.043-0.06mm) Merck. 5-Chloroisatin derivatives were synthesized by alkylation method available in the phase transfer catalysis conditions [6, 7]. The synthetic strategies adopted to obtain target compounds are depicted in Figure 1 and 2.

1.1 GENERAL PROCEDURE FOR THE SYNTHESIS OF NEW 5-CHLOROISATIN DERIVATES:

Compounds were then reacted with various alkylating agent, so into a 100 mL two-necked flask we introduce 0.2 g (1.1 mmol) of 5-chloro-1H-indole-2,3-dione (0.23 g, 1.16 mmol) of potassium carbonate in 15 ml of N-N-dimethylformamide (DMF) and (0.035 g, 0.10 mmol) of BTBA with magnetic stirring, the alkylating agent is added slowly, the mixture is left at room temperature for 48 hours. During this period the progress of the reaction is monitored by TLC (thin layer chromatography). Once the reaction is complete, the salts are removed

by filtration, the solvent (DMF) is evaporated under reduced pressure. The product obtained is purified

on a column of silica gel eluent (ethyl acetate / hexane).

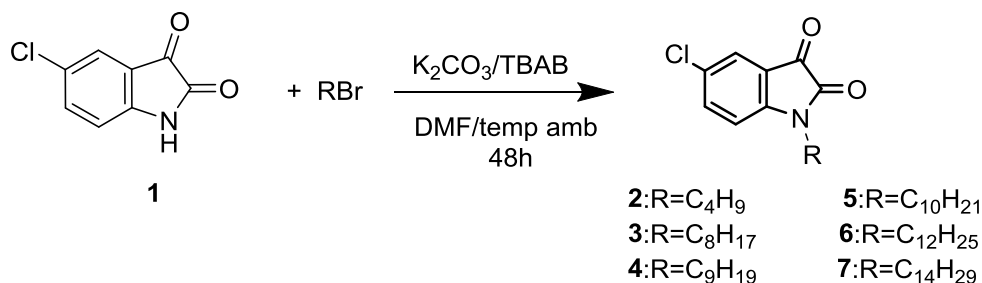


Figure 1: Synthesis of new products from 5-chloro-1H-indole -2,3-dione

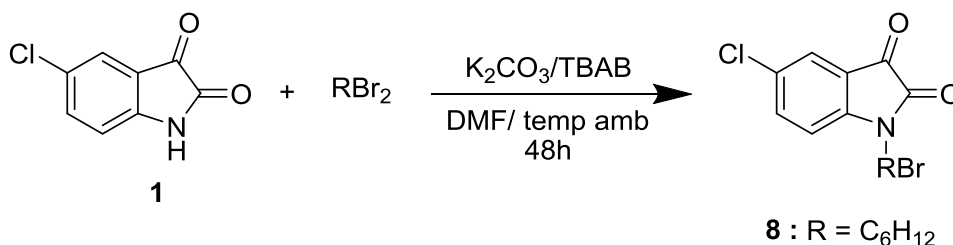


Figure 2: Synthesis of novel derivatives of 5-Chloro-1H-indole -2,3-dione by catalysis by phase transfer

1.2 SPECTRAL DATA

Compound 2 :1-butyl-5-chloroindoline-2,3-dione:

Yield: 85% ; m.p: 80-82°C; $R_f = 0.85$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.54-7.55 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) 7.50 7.51(d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 6.84 (d, H, H_{Ar} , $^3J_{H-H} = 6\text{Hz}$) ; 3.72 (t, 2H, CH_2 , $^3J_{H-H} = 6\text{Hz}$) ; 1.60-1.70 (m, 2H, CH_2) ; 1.35-1.42 (m, 2H, CH_2) , 0.95 (t, 3H, CH_3 , $^3J_{H-H} = 9\text{Hz}$). ^{13}C NMR (CDCl_3 ; 75MHz): δ (ppm) 183.83 (C=O) ; 163.28 (N-C=O) ; 142.43, 136.12, 121.06 (Cq); 137.61, 125.38, 111.40 (CH_{Ar}) ; 40.16, 29.24, 20.14 (CH_2) ; 13.67 (CH_3).

Compound 3 : 5-chloro-1-octylindoline-2,3-dione:

Yield: 88% ; m.p: 68-70°C ; $R_f = 0.85$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.54-7.55(d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 7.50-7.51 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 6.84 (d, H, H_{Ar} , $^3J_{H-H} = 9\text{Hz}$) ; 3.68 (t, 2H, CH_2 , $^3J_{H-H} = 6\text{Hz}$) ; 1.71-1.53 (m, 2H, CH_2) ; 1.24-1.31 (m, 10H, CH_2) , 0.85 (t, 3H, CH_3 , $^3J_{H-H} = 6\text{Hz}$). ^{13}C NMR (CDCl_3 ; 75MHz): δ (ppm) 183.20 (C=O) ; 160.73 (N-C=O) ; 144.60, 137.60, 125.37 (Cq); 129.45, 115.79, 111.42 (CH_{Ar}); 40.47, 31.73, 29.19, 27.20, 26.88, 22.60(CH_2) ; 14.04 (CH_3).

Compound 4 : 5-chloro-1-nonylindoline-2,3-dione:

Yield: 87% ; m.p: 67-68 °C; $R_f = 0.82$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.58-7.59 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 7.54-7.55 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 6.85 (d, H, H_{Ar} , $^3J_{H-H} = 9\text{Hz}$) ; 3.70 (t, 2H, CH_2 , $^3J_{H-H} = 9\text{Hz}$) ; 1.65-1.75(m, 2H, CH_2) ; 1.28 (m, 12H, CH_2) , 0.89 (t, 3H, CH_3 , $^3J_{H-H} = 6\text{Hz}$). ^{13}C NMR (CDCl_3 ; 75MHz): δ (ppm) 181.84 (C=O); 167.00 (N-C=O);

147.07, 137.63, 129.39 (Cq); 141.28, 116.10, 110.29 (CH_{Ar}); 42.53, 32.42, 29.49, 29.27, 29.21, 27.21, 26.89, 23.18 (CH_2); 15.83 (CH_3).

Compound 5: 5-chloro-1-decylindoline-2,3-dione:

Yield: 86% ; m.p: 62-64°C; $R_f = 0.77$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.53 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 7.50 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 6.80 (d, H, H_{Ar} , $^3J_{H-H} = 9\text{Hz}$) ; 3.67 (t, 2H, CH_2 , $^3J_{H-H} = 6\text{Hz}$) ; 1.60-1.67 (m, 2H, CH_2) ; 1.22 (m, 14H, CH_2) , 0.84 (t, 3H, CH_3 , $^3J_{H-H} = 6\text{Hz}$). ^{13}C NMR (CDCl_3 ; 75MHz): δ (ppm) 183.15 (C=O); 166.03 (N-C=O); 145.52, 129.39, 125.39 (Cq); 141.28, 115.58, 111.40 (CH_{Ar}); 40.49, 31.87, 29.49, 29.27, 29.21, 27.21, 26.89, 22.70 (CH_2) ; 14.12 (CH_3).

Compound 6 :5-chloro-1-dodecylindoline-2,3-dione:

Yield: 85% ; m.p: 64-66°C; $R_f = 0.75$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.55 (d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 7.51(d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 6.82 (d, H, H_{Ar} , $^3J_{H-H} = 9\text{Hz}$) ; 3.69 (t, 2H, CH_2 , $^3J_{H-H} = 9\text{Hz}$) ; 1.61-1.70(m, 2H, CH_2) ; 1.23(s, 18H, CH_2) , 0.86(t, 3H, CH_3 , $^3J_{H-H} = 6\text{Hz}$). ^{13}C NMR (CDCl_3 ; 75MHz): δ (ppm) 184.39 (C=O); 162.95 (N-C=O); 149.63, 129.47, 118.17(Cq); 137.64, 125.38, 111.44 (CH_{Ar}); 40.49, 31.93, 29.61, 29.45, 29.46, 29.34, 29.21, 27.21, 26.89, 22.70 (CH_2) ; 14.13 (CH_3).

Compound 7 : 5-chloro-1-tetradecylindoline-2,3-dione:

Yield: 84% ; m.p : 65-67°C ; $R_f = 0.73$; ^1H NMR (CDCl_3 ; 300MHz): δ (ppm) 7.54(d, H, H_{Ar} , $^4J_{H-H} = 3\text{Hz}$) ; 7.50 (d, H, H_{Ar} , $^4J_{H-H} = 2.1\text{Hz}$) ; 6.84 (d, H, H_{Ar} , $^3J_{H-H} = 9\text{Hz}$) ; 3.68 (t, 2H, CH_2 , $^3J_{H-H}$

=9Hz); 1.60-1.70 (m, 2H, CH₂); 1.22 (s, 22H, CH₂), 0.85 (t, 3H, CH₃, ³J_{H-H} = 6Hz). ¹³C NMR (CDCl₃; 75MHz): δ(ppm) 182.24 (C=O); 161.85 (N-C=O); 137.62, 125.35, 111.41(Cq) ; 130.45, 129.43, 118.46 (CH_{Ar}); 40.48, 31.94, 29.65, 29.62, 29.61, 29.54, 29.46, 29.36, 29.21, 27.20, 26.88, 22.70 (CH₂); 14.12 (CH₃).

Compound 8: 1-(6-bromoheptyl)-5-chloroindoline-2,3-dione:Yield: 85 % ; m.p: 66-70 °C ; R_f = 0.8 (Hexane/EtOAc, 2:1); ¹H NMR (CDCl₃; 300MHz): δ (ppm) 7.49-7.51 (dd, H, CH_{Arom}, J_{H-H} = 1.5 Hz, J_{H-H} = 4.5 Hz) ; 7.47 (d, H, CH_{Ar}, J_{H-H} = 1.5 Hz) ; 6.77 (d, H, CH_{Arom}, J_{H-H} = 6 Hz) ; 3.65 (t, 2H, CH₂, J_{H-H} = 3Hz) ; 3.33 (t, 2H, CH₂, J_{H-H} = 6Hz) ; 1.75-1.82 (m, 2H, CH₂), 1.61-1.68 (m, 2H, CH₂) , 1.47 (m, 4H, CH₂). ¹³C NMR (CDCl₃; 75MHz): δ (ppm): 182.64 (C=O); 160.28 (N-C=O); 146.49, 132.61, 119.49 (Cq) ; 134.44, 130.93, 123.33 (CH_{Ar}); 40.51 , 33.70 , 32.01, 27.89, 27.07, 26.15 (CH₂) .

II. IN VITRO ANTIBACTERIAL ACTIVITY EVALUATION

2.1 A disc diffusion test:

This method makes it possible to evaluate the antibacterial activity of a product. Although it is recognized as reliable and reproducible, it is mostly used as a preliminary stage for further studies as it provides access to essentially qualitative results. The technique used is a modification of the method of Hayes and Markovic.

It consists of depositing a sterile disk imbibed by the test product on a bacterial mat at the very beginning of its growth and measuring the area where the bacteria could not develop. The inhibition diameter, which reflects the antibacterial activity of the product tested, is thus determined. The antimicrobial activity of 5-Chloro-1*H*-indole-2,3-dione derivatives was evaluated using the disc diffusion method [8] using different microorganisms, including three types of bacteria: *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*. Mueller Hinton agar medium (MHA) was used for bacteria. Plates were preincubated at 37 °C for 24 h.

A sterile paper disk (6 mm in diameter) was placed on the surface of each agar plate and impregnated with 5 µL of each solution of 5-chloroisatin derivatives at a final concentration of 10 mg / ml. Then, the Petri dishes are incubated at 37 °C for 24 h for the bacteria. The diameters of the inhibition zones were measured in mm (including the diameter of the disk) with the caliper. A disc impregnated with 2% dimethylsulfoxide as a negative control was made. Each experiment was carried out in triplicate.

2.2 MINIMUM INHIBITORY

CONCENTRATION DETERMINATION (MIC):

The minimal inhibitory concentration (MIC) of our products was evaluated according to the microdilution method with some modifications. The minimum inhibitory concentration (MIC) was determined as the lowest concentration with no microbial growth compared to the positive control. DMSO (2%) was used as a negative control.

Briefly, the dilution of the 5-chloroisatin derivatives was prepared in a Mueller Hinton broth supplemented with bacteriological agar, to reach a final concentration between 5 mg / ml and 0.004 mg / ml, 50 µL of bacterial inoculum was added to each well at a final concentration of 106 CFU / ml.

The final concentration of our product was between 5 mg ml⁻¹ (3rd well) and 0.019 mg ml⁻¹ (well 11). Plates were incubated at 37°C for 24 h. After 2 hours of a subsequent incubation, bacterial growth was revealed by reduction of blue dye resazurin to pink resorufin [9].

2.3 MINIMUM BACTERICIDAL

CONCENTRATIONS (MBC):

order to determine the CMB, a bactericidal control is carried out 24 hours earlier by streaking on a platelet agar, after microdilution to the broth by spreading 5 µl of the negative wells on Luria Bertani agar plates (Luria Bertani medium: Yeast extract 5.0 g, peptone 10.0 g, sodium chloride 5.0 g, distilled water 1000 ml). The MBC will be the lowest concentration whose transplant shows a growth of germ less than or equal to 0.01% of survivors [10].

III. RESULTS AND DISCUSSION :

Table 1: MICs and MBCs of the compounds against the microbes used

compounds	MIC/MBC (mg/mL)			
	Gram+		Gram-	
	<i>Bacillus cereus</i>	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>
2	0,156/0,156	0,313/0,313	-	-
3	0,078/0,078	0,156/0,156	-	-
4	0,078/0,078	0,01/0,01	-	-
5	0,625/0,625	0,313/0,313	-	-

6	-	-	-	-
7	-	-	-	-
8	-	1,25/1,25	1,25/1,25	

This table showed that compound **4** exercised an excellent inhibitory activity against *Staphylococcus aureus* à Gram positive bacteria, with a MIC value of 0,01 mg/mL, while compounds **3** and **4** displayed comparable activity against *Bacillus cereus* with a MIC value of 0,078 mg/ml, while compounds **3** and **5** displayed moderate to good activity against Gram positive bacteria studied. The compounds **8** exhibited moderate inhibitory effect against Gram+ and Gram- bacteria (*staphylococcus aureus* and *Escherichia coli*) at a MIC value of 1,25 mg/mL. However, *Pseudomonas aeruginosa* (à Gram – bacteria) was resistant to all tested compounds. Moreover, the compounds **6** and **7** did not show any antimicrobial effects against all tested strains. Generally, Gram-positive bacteria are much more susceptible to antibacterial agents than Gram negative bacteria, whose resistance is attributed to the structures of their cell wall. Gram-negative bacteria have a thick lipid bilayer which is selectively permeable [11].

IV. CONCLUSION

The synthesis of a variety of the 5-Chloroisatin derivatives was carried out via the N-alkylation reaction. All compounds were obtained in good yields. The structures of compounds obtained were confirmed by ¹H-NMR, ¹³C-NMR. The antimicrobial activity of these compounds was tested against two Gram-negative bacteria and two Gram-positive bacteria. From the above results it can be concluded that, the highest activity was obtained with the **4** compound which exhibited good to high activity towards *Staphylococcus aureus* - Gram positive bacteria, with a MIC value of 0.01 mg / ml. Compounds **2**, **3**, **5**, **8** showed moderate activity against the Gram positive bacteria studied, whereas compounds **6** and **7** showed no antimicrobial effect.

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